# The impact of oestrus synchronization by hormone medication and varying numbers of artificial insemination sessions, determined by sow reproductive indices

Jani Mavromati\* and Luigj Turmalaj

### **Summary**

This study was conducted on a central Albanian pig farm with a capacity of 80 sows. A uniform distribution of piglets born and the number of reared piglets sold every week and month was the target. To achieve this goal, a hormone-based synchronization scheme was used, combined with the use of one, two, or three artificial insemination sessions during the same oestrus. The main reproduction indices of sows were monitored, and the results indicated that the farrowing rate after the first insemination was 78.57%, 83.64% and 83.33% for the three groups in which one, two and three artificial insemination sessions were used during the same oestrus, obtained by inoculation with the hormone Sergon PG 400/200 IU. Average litter size (the number of live born piglets) per farrowing sow was  $10.22 \pm 0.33$ ,  $10.8 \pm 0.2$  and  $10.6 \pm 0.3$  in the three groups, respectively. The fecundity index (FI) was 803, 905.8 and 883.3 in the three experimental groups of sows, respectively. A total of 1436 piglets from 136 farrowing sows were sold during 2018, achieving uniform sales of over 100 pigs per month with a weight over 90 kg. Therefore, it can be concluded that oestrus synchronization of sows with the Sergon PG 400/200 hormone can achieve a uniform distribution of piglets born and consequently a uniform distribution of piglets sales per week or month, regardless of the fact that this should be achieved at a larger scale. The best result as seen in the reproductive indicators was achieved by the use of artificial insemination with two sperm inoculations at a 12-hour interval during the same oestrus, synchronized by the Sergon PG 400/200 hormone.

**Key words**: artificial insemination; oestrus synchronization; hormone; reproduction indexes; sows

Jani MAVROMATI, DVM, PhD, (Corresponding author, e-mail: e-mail: jmavromati@ubt.edu.al), Assistant Professor, Department of Veterinary Public health, Faculty of Veterinary Medicine, Agricultural University of Tirana, Albania; Luigj TURMALAJ, DVM, PhD, Full Professor, Department of Veterinary clinics, Faculty of Veterinary Medicine, Agricultural University of Tirana, Albania

### Introduction

Pork producers in Albania are interested in increasing reproductive performance on their farms. Greater control and distribution of born piglets is implemented with the aim of better planning of monthly sales. The use of hormonal drugs for the synchronization of oestrus and consequently piglets, and the use of biotechnology for the artificial insemination of sows, remain challenges for farmers in this sector. The advantages of artificial insemination in sows are numerous. There are different techniques available, such as intra cervical insemination (Cassar et al., 2005; Caravaca et al., 2012), Post-Cervical Insemination (PCI; Rath et al., 2000; Knox et al., 2011; Fontana et al., 2014) and Deep intrauterine insemination (DIU), or intra uterine horns techniques (Watson et al., 2001; Dimitrow et al., 2007).

Different factors influence the fertility of liquid boar semen, including storage time, conservation, season and the constitutions of dilutions (Koh et al., 1976; Sulo, 1985; Alexopoulos et al., 1996; Roca et al., 2006).

Different techniques use different volumes of liquid inseminate, and in turn different numbers of spermatozoa per dose (Wiggins et al., 1951; Krueger et al., 1999; Martinez et al., 2001).

The target of these techniques is the number of doses collected per boar during the year. The number of boars on the farm improve the quality of semen and quality of boars (Table 1).

According to the literature, the number of spermatozoa per dose for artificial insemination is very important. In cervical procedures of the artificial insemination (CAI) in sows, billions of spermatozoa are used (2.5-4 x 109 spermatozoa per insemination dose) in varying volumes of liquid per dose (70-100 mL). Approximately 90% of the spermatozoa inseminated cannot be recovered from the uterus 2 hours after artificial insemination (AI) (Pursel et al., 1978; Viring, 1980). According to Mburu et al. (1996), only 1 x 10<sup>5</sup> spermatozoa reach the uterotubal junction and about 1 x 103 reach the sperm reservoir in the caudal 1-2 cm of the isthmus (Hunter, 1981),

Table 1. Different techniques of artificial insemination in sows, their volume and number of spermatozoa

	Traditional artificial insemination Intra Cervical (CIA)	Post Cervical (PCI)	Intra Uterine horn insemination or Deep intrauterine (DIU)
Number of Spermatozoa	(3x10 <sup>9</sup> /80-100 mL)	(1.5x10°/40-50 mL)	(0.6-1x10°/20-30 mL)
Volume of liquid	100 mL	50 mL	20 mL
Frequency of Ejaculation per boar/week	2 times/week	2 times/week	2 times/week
Doses per ejaculation	20	40	100
Doses per year	2,000	4,000	10,000
Proportion of boars	10	5	2

Table adapted from the literature

where the cells can be stored without reducing their fertilizing ability (Suarez et al., 1991) until just before ovulation (Hunter, 1984). Sufficient spermatozoa to ensure subsequent fertilization are established in the isthmus reservoir within one hour of mating (Hunter, 1981, 1984). Some of the principal reasons for loss of spermatozoa after insemination are backflow (Steverink et al., 1998; Matthijs et al., 2003), phagocytosis by polymorphonuclear neutrophils (PMN), which influx the porcine uterus after insemination (Rozeboom et al., 1999; Matthijs et al., 2003), and sperm losses in the uterine horns caused by the adhesion of sperm to the ciliary epithelial cells of the uterus (Levis et al., 2001).

Other studies have investigated the use of one, two or three artificial insemination sessions during the same oestrus. The number of insemination sessions can affect the cost of insemination (Alexopullos et al., 1996; Caravaca et al., 2012; Driancourt et al., 2013; Ulguim et al., 2016).

Depending on the situation, the most effective technique for the best fecundity index should be selected at the individual farm level, and this depends on farrowing rate and the number of piglets born alive per litter. For this reason, different hormones have been used in the past to stimulate oestrus synchronization (Cox et al., 1983; Dial and BeVier, 1986; Britt et al., 1989). Based on the above techniques, a new group of hormones is used in protocols for oestrus synchronization (De Rensis and Kirkwood, 2003; Baer and Bilkei, 2004; Degenstein et al., 2008; Manjarin et al., 2010; Martinat-Botté et al., 2010; de Jong et al., 2013; Ulguim et al., 2016).

The purpose of this study was to assess the differences in reproductive parameters in sows following artificial insemination with one, two or three sperm inoculations during the same oestrus, synchronized using the Sergon PG 400/200 hormonal drugs produced by the Bioveta Cz company (lyophilized

powder of two hormonal substances: human chorionic gonadotropin (hCG) and equine chorionic gonadotropin (eCG)). Sergon PG 400/200 was applied in accordance with the manufacturer's recommendations (Bioveta, 2018).

The study was performed using the traditional artificial insemination technique, Cervical artificial insemination (CAI) in sows in the most effective and economical way, to achieve uniform sales of rearing pigs for each week or month associated with oestrus synchronization through hormonal treatment.

### **Material and Methods**

The study was conducted on a pig farm in central Albania with a capacity of 80 sows during 2018. The aim of the study was to evaluate the fecundity index (FI) which depends on the farrowing rate and the number of piglets born alive per litter, by insemination with one, two, or three artificial insemination sessions during the same oestrus, synchronized by the Sergon PG 400/200 hormone (Bioveta, Czech Republic; Bioveta, 2018).

### Hormonal treatment

The second day after piglets were born, sows were injected with 2 mL I/M Sergon PG 400/200 IU. Oestrus was detected 2-3 days later. Despite the appearance of oestrus, sows were not fertilized at this time. At 21 days after oestrus, artificial insemination was performed. Sows that did not respond after the first treatment were subjected to a second treatment over a 25-day period with the same hormone (Sergon PG 400/200 IU, 2 mL I/M). Sows that did not display oestrus after the second treatment were excluded from the study (Bioveta, 2018).

Serum gonadotropin acts similarly to follicle stimulating hormone (FSH) and the luteinizing hormone (LH) and induces the growth of ovarian follicles. Chorionic gonadotropin acts similar to

LH and supports ovulation and corpus luteum growth. The combination of these hormones induces the fertile oestrus cycle in sows (Bioveta, 2018).

### **Sperm collection**

The same boar was used for the insemination of the three groups of sows (large white), Sperm was diluted with BTS (Beltsville Thawing Solution, Minitüb, Germany).

The boar was used to obtain semen once a week using the gloved-hand technique and semen was filtered to remove the gel. The average number of spermatozoa/AI dose was controlled by visual estimation of density according to colour, and measured with a colorimeter. Semen quality assessment, including viability, motility, progressive motility and morphology of spermatozoa, was estimated using a bipolar microscope. After calculation and dilution, doses were stored in 100 mL bottles containing 3 billion (3×109) spermatozoa per dose. Doses were stored at 16°C and used within 24 h, according to Levis et al. (2001). CAI was performed with disposable spiral tip catheters (Minitüb, Germany).

### Sperm inoculation

Sergon PG 400/200 IU was inoculated into 166 large white sows after farrowing for the synchronization of oestrus. Oestrus detection was performed twice daily by experienced workers and by allowing sows nose-to-nose contact with mature boars and applying backpressure. The occurrence of oestrus was defined by

the standing reflex in front of a teaser boar and reddening and swelling of the vulva, according Levis et al. (2001).

After thorough cleaning and drying the sow's vulvar labia area, the insemination dose was introduced slowly in the sow's uterus. The catheter remained in the cervix 2–4 min after insemination to reduce backflow, according to Levis et al. (2001).

Of the 166 sows, 56, 62 and 48 were artificially inseminated with one, two, or three inoculations of sperm, respectively. The age of the sows in the three groups did not differ significantly (P > 0.1) with respect to the number of gestations. Other breeding conditions, such as nutrition, prophylactic programmes, etc., were the same for all three groups of sows.

In the first group in which only one dose of semen was inoculated, insemination was performed 12 hours after the discovery of oestrus when the sow stayed with the boar. For the second and third doses, the first insemination was carried out 12 hour after the discovery of oestrus, and the second or third insemination was performed at 12-hour intervals after the first insemination.

### **Results and discussion**

The study was conducted to assess the fertility parameters and the total number of piglets born alive per sow, after the insemination of 166 sows with one, two or three insemination sessions during the same oestrus following hormone synchronization.

<b>Table 2.</b> Number of sows for each group and the hormonal medication use	ed
-------------------------------------------------------------------------------	----

Groups	No. of sows	Number of inseminations during the same oestrus	Hormone medication used for oestrus synchronization
Group A	56	1	Sergon PG 400/200 IU
Group B	62	2	Sergon PG 400/200 IU
Group C	48	3	Sergon PG 400/200 IU

Utjecaj sinkronizacije estrusa hormonalnim lijekovima i različitim brojem umjetnih osjemenjivanja u reproduktivnim indeksima krmača

Table 3. Fertility after the first insemination session for three groups of sows

Groups	No. of sows	Number of inseminations during the same oestrus	Farrowing on the first oestrus	Hormone medication used for oestrus synchronization
Group A	56	1	44 sows (78.57%)	Sergon PG 400/200 IU
Group B	62	2	52 sows (83.87%)	Sergon PG 400/200 IU
Group C	48	3	40 sows (83.33%)	Sergon PG 400/200 IU

Table 4. Number of piglets born live for each group and for each litter

Groups	No. of sows	Number of inseminations during the same oestrus	Live born litter size
Group A	56	1	10.22±0.3 per sow
Group B	62	2	10.8±0.2 per sow
Group C	48	3	10.6±0.3 per sow

Table 5. Summary table for fertility and number of piglets born live per sow in three groups

Groups	No. of sows	Number of inseminations during the same oestrus	Farrowing on the first oestrus	Live born litter size	Hormone medication used for oestrus synchronization
Group A	56	1	44 sows (78.57%)	450 piglets born (10.22±0.3/litter)	Sergon PG 400/200 IU
Group B	62	2	52 sows (83.87%)	562 piglets born (10.8±0.2/litter)	Sergon PG 400/200 IU
Group C	48	3	40 sows (83.33%)	424 piglets born (10.6±0.3/litter)	Sergon PG 400/200 IU

The table above shows the percentage of fertility in three groups of artificial insemination of sows receiving one, two or three semen inoculations during the same oestrus, following synchronisation using the hormone Sergon PG 400/200 IU.

The percentage of sow fertility after artificial insemination during the first oestrus was 78.57%, 83.64 and 83.33% for the three groups, respectively, receiving one, two or three semen inoculation during the same oestrus, and the use of the same hormone.

The number of piglets born live per litter was  $10.22 \pm 0.33$ ,  $10.8 \pm 0.2$  and  $10.6 \pm 0.3$  piglets for the three groups of sows, respectively. In the second and third group of sows, there was no difference in the fertility percentage, or in the average number of piglets born live per litter. These results are consistent with other reports (Britt et al., 1989; Bates et al., 1991).

However, in the first group, which received artificial insemination only once during the same oestrus, a difference of

	Cervical Artificial Insemination (CAI)			
	CAI-1 insemination 12 hours after oestrus (Group 1)	CAI-2 insemination12 and 24 hours after oestrus (Group 2)	CAI-3 insemination12, 24, 36 hours after oestrus (Group 3)	
No. of sows (n)	56	62	48	
Farrowing rate (%)	78.57 % ( <i>n</i> =44)	83.87 % ( <i>n</i> =52)	83.33% ( <i>n</i> =40)	
Return, abortion, etc. (%)	21.43% (n=12)	16.13 ( <i>n</i> =10)	16.7% ( <i>n</i> =8)	
Live born litter size (n)	10.22±0.3	10.8±0.2	10.6±0.3	
Fecundity index - FI (n)	803	905.8	883.3	
Hormone for oestrus synchronization	Sergon PG 400/2001U	Sergon PG 400/200 IU	Sergon PG 400/200 IU	

**Table 6.** Reproductive parameters obtained for three groups of sows inseminated by CAI, with one, two and three inseminations in the same oestrus, with the use of the hormone Sergon PG 400/200 IU

about 5% was observed in the farrowing rate (78.57%), with a difference of approximately 0.5 piglets on average per litter, less than in the second and third groups of sows (Tables 5 and 6).

According to some authors, the percentage of fertility in artificial insemination may (Roca et al., 2006) or may not be influenced (Sulo, 1985) by the number of sperm inoculations within the same oestrus. This can also be influenced by other factors such as number of spermatozoa per sperm dosage (Alexopuolos et al., 1996), breeder, environmental factors (Koh et al., 1976), and so on. Another important factor that can influence artificial insemination is also the time of oestrus appearance and correct timing of sperm inoculation (Alm et al., 2006).

### Distribution of born piglets and number of rearing pigs on the market each month

From the tables above, we can see that in the year 2018, 136 fertilized sows delivered a total of 1436 piglets (Table 5, groups a, b, c). Over 1200 rearing pigs (*i.e.* over 100 rearing pigs per month) were sold during 2018, with a weight of

over 90 kg per pig at age near 5 months. Therefore, over 100 rearing pigs per month were obtained (thereby fulfilling the main request from the farm).

## Conclusions and Recommendations

The results of this study and findings from the literature allow us to conclude that the percentage of sow fertility and total number of piglets born on average per sow using artificial insemination with one, two or three inoculations during the same following synchronization oestrus using hormonal drugs (Sergon PG 400/200 IU) have only inconspicuous differences. The best results were obtained using insemination with 2 or 3 sperm inoculations than with a single inoculation during the same oestrus synchronized by hormonal drugs.

Specifically, the results of sow fertility were 78.57%, 83.64 and 83.33% for the three groups, respectively, in which one, two and three inoculations were used during the same oestrus induced by the hormonal drugs like Sergon PG 400/200 IU.

The total number of piglets born live was  $10.22 \pm 0.33$ ,  $10.8 \pm 0.2$  and  $10.6 \pm 0.3$  per litter for the three groups, respectively. The second and third group of sows showed no differences in fertility or in the number of piglets born live per litter.

In the first group, receiving artificial insemination only once during oestrus, fertility was about 5% percent lower (78.57%) with a margin of approximately 0.5 piglets per litter less than in the second and third groups.

Based on the results of this study, we recommend pig farmers should use artificial insemination with two sperm inoculations during the same oestrus, following hormonal synchronization using Sergon PG 400/200 IU.

Using only a single inoculation can give lower results. The use of three separate inoculations, however, is not economical and had no effect on increasing the reproductive index in sows.

Thus, the greatest interest in practical use in terms of fertility percentage in sows, the total number of piglets born live per litter and the economic benefit was achieved by administering two sperm inoculations in a 12-hour interval in the same oestrus, following hormonal synchronization with Sergon PG 400/200 IU.

### References

- ALEXOPOULOS, C., C. BOSCOS, PH. SARATSIS, C. SAULIDIS and S. KYRIAKIS (1996): The effect of storage time and number of spermatozoa per insemination dose on semen characteristics and fertilizing capacity of boar semen diluted with Beltville Thawing Solution (BTS) extender. Anim. Sci. 62, 599-604.
- ALM, K., O. PELTONIEMI, E. KOSKINEN and M. ANDERSSON (2006): Porcine Field Fertility with Two Different Insemination Doses and the Effect of Sperm Morphology. Reprod. Domest. Anim. 41, 210-213.
- BATES, R. O., B. N. DAY, J. H. BRITT, L. K. CLARK and M. A. BRAUER (1991): Reproductive performance of sows treated with a combination of

- pregnant mare's serum gonadotropin and human chorionic gonadotropin at weaning in the summer. J. Anim. Sci. 69, 894-898.
- 4. Bioveta Animal Health (2018): www.bioveta.eu
- BRITT, J. H., B. N. DAY, S. K. WEBEL and M. A. BRAUER (1989): Induction of fertile estrus in prepuberal gilts by treatment with a combination of pregnant mare's serum gonadotropin and human chorionic gonadotropin. J. Anim. Sci. 67, 1148-1153.
- BAER, C. and G. BILKEI (2004): The effect of intravaginal applied GnRH-agonist on the time of ovulation and subsequent reproductive performance of weaned multiparous sows. Reprod. Domest. Anim. 39, 293-297.
- CARAVACA, I. H., M. J. IZQUIERDO-RICO and F. A. GARCÍA-VÁZQUEZ (2012): Reproductive performance and backflow study in cervical and post-cervical artificial insemination in sows. Anim. Reprod. Sci. 136, 14-22.
- CASSAR, G, R., N. KIRKWOOD, Z. POLJAK, K. BENNETT-STEWARD and R. M. FRIENDSHIP (2005): Effect of single or double insemination on fertility of sows bred at an induced estrus and ovulation. J. Swine Health Prod. 13, 254-258.
- COX, N. M., K. L. ESBENSHADE and J. H. BRITT (1983): Treatment of long-term anestrous sows with estradiol benzoate and GnRH: response of serum LH and occurrence of estrus. Theriogenology 20, 499-507.
- De RENSIS, F. and R. N. KIRKWOOD (2016): Control of estrus and ovulation: Fertility to timed insemination of gilts and sows. Theriogenology 86, 1460-1466.
- DEGENSTEIN, K. L., R. O' DONOGHUE, J. L. PATTERSON, E. BELTRANENA, D. J. AMBROSE, G. R. FOXCROFT and M. K. DYCK (2008): Synchronization of ovulation in cyclic gilts with porcine luteinizing hormone (pLH) and its effects on reproductive function. Theriogenology 70, 1075-1085.
- DIAL, G. D. and G. W. BeVIER (1986): Pharmacologic control of estrus and ovulation in the pig. In: D. A. Morrow (ed.) Current Therapy in Theriogenology. 2<sup>nd</sup> ed. W. B. Saunders Company, Philadelphia, PA., pp. 912-914.
- de JONG E, J. KAUFFOLD, S. ENGL, J. JOURQUIN and D. MAES (2013): Effect of a GnRH analogue (Maprelin) on the reproductive performance of gilts and sows. Theriogenology 80, 870-877.
- DRIANCOURT, M. A., P. COX, S. RUBION, G. HARNOIS-MILON, B. KEMP and N. M. SOEDE (2013): Induction of an LH surge and ovulation by buserelin (as Receptal) allows breeding of weaned sows with a single fixed-time insemination. Theriogenology 80, 391-399.
- DIMITROW, S., E. JELIAZKOV and D. LEVIS (2007): Deep intrauterine and transcervical insemination of sows and gilts. Trakia J. Sci. 5, 40-46.
- FONTANA, D. L, R. R. ULGUIM, P. E. SBARDELLA, M. L. BERNARDI, I. WENTZ, F. P. BORTOLOZZO (2014): Fixed-time post-cervical

- artificial insemination in sows receiving porcine luteinising hormone at oestrus onset Anim. Reprod. Sci. 144, 109-114.
- HUNTER, R. H. F. (1981): Sperm transport and reservoirs in the pig oviduct in relation to the time of ovulation J. Reprod. Fertil. 63, 109-117.
- HUNTER, R. H. F. (1984): Pre-ovulatory arrest and peri-ovulatory redistribution of competent spermatozoa in the isthmus of the pig oviduct J. Reprod. Fertil. 72, 203-211.
- KOH, T. J., B. G. CRABO, H. L. TSOU and E. F. GRAHAM (1976): Fertility of liquid boar semen as influenced by breed and season. J. Anim. Sci. 42, 138-144.
- KNOX, R. V., K. L. WILLENBURG, S. L. RODRIGUEZ-ZAS, D. L GREGER, H. D. HAFS and M. E. SWANSON (2011): Synchronization of ovulation and fertility in weaned sows treated with intravaginaltriptorelin is influenced by timing of administration and follicle size. Theriogenology 75, 308-319.
- KRUEGER, C., D. RATH and L. A. JOHNSON (1999): Low dose insemination in synchronized gilts. Theriogenology 52, 1363-1373.
- LEVIS, D. G., S. BURROUGHS and S. WILLIAMS (2001): Use of Intra-Uterine Insemination of Pigs: Pros, Cons & Economics. University of Nebraska -Lincoln Digital Commons@University of Nebraska -Lincoln Faculty Papers and Publications in Animal Science Animal Science Department.
- MARTINAT-BOTTÉ, F., E. VENTURI, P. GUILLOUE, M. A. DRIANCOURT and M. TERQUI (2010): Induction and synchronization of ovulations of nulliparous and multiparous sows with an injection of gonadotropin-releasing hormone agonist (Receptal). Theriogenology 73, 332-342.
- MARTINEZ, E. A., J. M. VAZQUEZ, J. ROCA, X. LUCAS, M. A. GIL, I. PARRILLA and J. L. VAZQUEZ (2001): Successful non-surgical deep intrauterine insemination with small numbers of spermatozoa in sows. Reproduction 122, 289-296.
- MBURU, J. N., S. EINARSSON, N. LUNDEHEIM and H. RODRIGUEZ-MARTINEZ (1996): Distribution, number and membrane integrity of spermatozoa in the pig oviduct in relation to spontaneous ovulation Anim. Reprod. Sci. 45 109-121.
- MATTHIJS, A., B. ENGEL and H. WOELDERS (2003): Neutrophil recruitment and phagocytosis of boar spermatozoa after artificial insemination of sows, and the effects of inseminate volume, sperm dose and specific additives in the extender. Reproduction 125, 357-367.
- 27. MANJARIN, R., J. C. GARCIA, J. C. DOMINGUEZ, M. J. CASTRO, B. ALEGRE, J. D. MUNOZ and R.

- N. KIRKWOOD (2010): Effect of gonadotropin treatment on estrus, ovulation, and litter size in weaned and anestrous sows. J. Anim. Sci. 88, 2356-2360
- PURSEL, V. G, L. L. SCHULMAN and L. A. JOHNSON (1978): Distribution and morphology of fresh and frozen–thawed sperm in the reproductive tract of gilts after artificial insemination Biol. Reprod. 19, 69-76.
- RATH, D., C. KRUEGER and L. A. JOHNSON (2000): Low dose insemination technique in the pig. In: Boar Semen Preservation IV (Editors: L. A. Johnson and H. D. Guthrie), Allen Press, Inc., Lawrence, KS, pp. 115-118.
- ROZEBOOM, K. J., M. H. TROEDSSON, T. W. MOLITOR and B. G. CRABO (1999): The effect of spermatozoa and seminal plasma on leukocyte migration into the uterus of gilts. J. Anim. Sci. 77, 2201-2206.
- ROCA, J., J. VÁZQUEZ, M. GIL, C. CUELLO, I. A. PARRILLA and E. MARTÍNEZ (2006): Challenges in Pig Artificial Insemination. Reprod. Domest. Anim. 41, 43-53.
- SULO, X. H. (1985): Vlerësimi i tre holluesve të spermës së harçit në pjellorinë e dosave". Buletini i Shkencave Zooteknike dhe Veterinare. Nr4, 83-92.
- STEVERINK, D., N. SOEDE, E. BOUWMAN and B. KEMP (1998): Semen backflow after insemination and its effect on fertilization results in sows. Anim. Reprod. Sci. 54, 109-119.
- SUAREZ, S., K. REDFERN, P. RAYNOR, F. MARTIN and D. M. PHILLIPS (1991): Attachment of boar sperm to mucosal explants of oviduct in vitro: possible role in formation of a sperm reservoir Biol. Reprod. 44, 998-1004.
- ULGUIM, R. R., D. L. FONTANA, M. L. BERNARDI, I. WENTZ and F. P. BORTOLOZZO (2016): Single fixed-time artificial insemination in gilts and weaned sows using pLH at estrus onset administered through vulvar submucosal route. Theriogenology 86, 1072-1080.
- VIRING, S. (1980): Distribution of live and dead spermatozoa in the genital tract of gilts at different times after insemination Acta Vet. Scand. 21, 145-149.
- 37. WATSON, P. F., J. BEHAN, G. DECUADO-HANSEN and B. CASSOU (2001): Deep insemination of sows with reduced sperm numbers does not compromise fertility: A commercially-based field trial. Proceeding 6th International Conference on Pig Reproduction, University of Missouri, June 3-6. Page 135 (Abstract).
- WIGGINS, E. L., R. H. GRUMMER and L. E. CASIDA (1951): Minimal volume of semen and number of sperm for fertility in artificial insemination in swine. J. Anim. Sci. 10, 138-143.

# Utjecaj sinkronizacije estrusa hormonalnim lijekovima i različitim brojem umjetnih osjemenjivanja u reproduktivnim indeksima krmača

Dr. sc. Jani MAVROMATI, dr. med. vet., docent, Katedra za javno zdravstvo veterine, Veterinarski fakultet, Poljoprivredno sveučilište Tirana, Albanija; Luigj TURMALAJ, dr. med. vet., redoviti profesor, Odjel za veterinarske klinike, Veterinarski fakultet, Poljoprivredno sveučilište Tirana, Albanija

Ova studija provedena je na farmi svinja s kapacitetom od 80 krmača, koja se nalazi u središnjoj Albaniji. Cilj je bila ujednačena raspodjela oprasenih i broja prasadi za uzgoj prodavanih svaki tjedan i mjesec. Za postizanje tog cilja, rabljena je shema sinkronizacije na bazi hormona u kombinaciji s uporabom jednog, dva i tri umjetna osjemenjivanja tijekom istog estrusa. Iz praćenja glavnih reproduktivnih indeksa krmača nakon pripremljene sheme rezultati su pokazali da je stopa prasenja nakon prve oplodnje bila 78,57 %, 83,64 % i 83,33 % za tri skupine u kojima su rabljene jedno, dva, odnosno tri umjetna osjemenjivanja tijekom postignutog istog estrusa injekcijama hormona Sergon PG 400/200 IJ. Veličina okota (broj živorođene prasadi) bila je 10,22  $\pm 0.33$ ,  $10.8 \pm 0.2$ , odnosno  $10.6 \pm 0.3$  prosječno rođene prasadi po oprasenim krmačama u tri skupine. Indeks plodnosti (FI) bio je 803; 905,8, odnosno 883,3 u tri eksperimentalne skupine krmača. Prodano je 1436 prasadi od 136 oprasenih krmača tijekom 2018., postigavši ujednačenu raspodjelu prodaje od više od 100 prasadi u mjesec, težine veće od 90 kg. Iz naših rezultata možemo zaključiti (bez obzira na činjenicu da je iste potrebno analizirati na većem broju grla) da se sinkronizacijom estrusa u svinja pomoću hormona Sergon PG 400/200 može postići ujednačena raspodjela oprasenih prasadi time ujednačena raspodjela prodaje prasadi tjedno i mjesečno. Najveći uspjeh u reproduktivnim indikatorima pokazala je uporaba umjetnog osjemenjivanja s dvije inokulacije sperme u razdoblju od 12 sati tijekom istog estrusa sinkroniziranog s hormonom Sergon PG 400/200.

Ključne riječi: umjetno osjemenjivanje, sinkronizacija estrusa, hormon, reproduktivni indeksi, krmače